198. Synthesis, Structure, and Antimalarial Activity of Tricyclic 1,2,4-Trioxanes Related to Artemisinin

by Charles W. Jefford*, Javier A. Velarde, Gérald Bernardinelli^a), Dorothy H. Bray, David C. Warhurst^b), and Wilbur K. Milhous^c)

 ^a) Department of Organic Chemistry, University of Geneva, CH-1211 Geneva 4
 ^b) PHLS Malaria Reference Laboratory, London School of Hygiene & Tropical Medicine, Keppel Street, London WC1E 7HT, UK
 ^c) Division of Experimental Therapeutics, Walter Reed Army Institute of Research, Washington, D.C. 20307-5100, USA

(3. VIII.93)

Two sets of tricyclic 1,2,4-trioxanes containing the ABC (10, 11) and ACD ring portions (21, 22, 32, 33, 37, and 38) of artemisinin (1) were synthesized by successive photo-oxygenation of appropriate enol-ether precursors to 1,2-dioxanes and inter- and intramolecular reaction with a carbonyl compound or oxo-substituted side-chain. The structures of 10, 21, and 22 were determined by X-ray analysis. The anti-malarial activity of all trioxanes, except 37 and 38, was evaluated *in vitro* against chloroquine-sensitive and chloroquine-resistant *Plasmodium falciparum* parasites. Trioxanes 11 and 21 were as active as artemisinin (1). It was found that neither the lactone function nor rings B and D of 1 were essential for activity. A possible pharmacophore for artemisinin-like activity, which embodies a spirocyclopentane group attached to C(3) of 1,2,4-trioxane, was proposed.

Introduction. – The worldwide spread of the resistance of *Plasmodium falciparum* to existing antimalarial agents together with the incidence of cross-resistance to structurally related derivatives makes the search for new drugs a matter of high priority [1]. The discovery that a naturally occurring sesquiterpenic 1,2,4-trioxane, artemisinin (1), is endowed with potent antimalarial properties has provided a providential lead for the development of improved versatile analogues [2]. However, this opportunity has not been exploited so far. Instead, derivatives of 1, namely arteether (2), artemether (3), sodium artesunate (4), and sodium artelinate (5) were chosen for clinical trials, despite difficulties in administration arising from poor solubility and stability [3] [4]. It occurred to us that the first step in designing better drugs based on 1 was to determine which parts of the skeleton were essential for the high activity characteristic of the parent structure. It is



1



2 R = Et **3** R = Me **4** R = NaO₂C(CH₂)₂C(O) **5** R = 4-NaO₂C-C₆H₄CH₂



already known that the endoperoxide bridge in 1 is a key structural element, because the metabolite **6**, where only an ether bridge remains, is devoid of activity [5]. Moreover, recent studies on cyclic peroxides and tetrahydrobenzopyran-derived 1,2,4-trioxanes demonstrated that neither the peroxide function nor the 1,2,4-trioxane ring alone are sufficient to confer significant antimalarial activity [6]. Consequently, in an attempt towards defining the structure-activity relationship, we synthesized several 1,2,4-trioxanes structurally resembling artemisinin and hereby report on their antimalarial activity *in vitro* against *Plasmodium falciparum*.

Chemistry. – Artemisinin (1) is a tetracyclic molecule composed of a cyclohexane, a δ -lactone, a 1,2,4-trioxane, and a 1,2-dioxepane moiety designated A, B, C, and D, respectively. We decided to synthesize two sets of tricyclic artemisinin-like compounds which embody just three of these cyclic features.

The ABC ring portion of artemisinin (1) was constructed by taking (-)-isopulegol (7) as the homochiral starting material or A ring component (*Scheme 1*). The B ring was assembled in five steps to give dihydropyran derivative 8 [7]. Photo-oxygenation of the double bond was face-specific. The resulting exo-1,2-dioxetane 9 was then enlarged into the two 1,2,4-trioxanes 10 and 11 by the inclusion of acetone and cyclopentanone, respectively, on catalysis with trimethylsilyl trifluoromethanesulfonate (Me₃SiOTf). The



structures of 10 and 11 embody the precise configurational features of artemisinin (1) as the ABC rings possess the same chirality. However, the X-ray analysis of 10 showed that the trioxane ring, unlike that in 1, adopts a chair conformation in the solid state. There may be a conformational bias towards a boat in the corresponding spirocyclopentane derivative 11.

To prepare derivatives containing the ACD ring, an intramolecular variant of the above-mentioned process was undertaken. The point of departure was 3-(2-oxocyclo-hexyl)propanenitrile (12; *Scheme 2*)¹). Application of the *Horner-Wittig* reaction to 12 by using diphenyl(methoxymethyl)phosphine oxide [8] afforded the (E)- and (Z)-isomers 13 and 14, respectively, in a 1:1 ratio. After chromatographic separation, the identity of each was established by recourse to ¹H- and ¹³C-NMR spectroscopy.

Each isomer can adopt one of two chair conformations in which the side chain occupies either an equatorial (e) or an axial (a) position. The (E)-isomer 13 prefers the former, while the (Z)-isomer 14 favors the latter owing to the operation of $A^{1,3}$ strain [9]. Accordingly, the (E)- and (Z)-isomers are readily distinguishable from each other by the characteristic chemical-shift differences of C(2), C(6), and the attached H-atoms (*Table 1*). The proton

¹) For the sake of clarity, racemic mixtures are represented as single enantiomers in *Schemes* 2-7.





Table 1. ¹³C- and ¹H-NMR Chemical Shifts (δ in ppm) of compounds 13 and 14 in CDCl₃



contiguous to the MeO group is deshielded relative to the same proton in the other isomer [10]; this is the H_e -C(6) (Δ 0.54 ppm) in either the equatorial or axial conformation of 13 and the H_e -C(2) (Δ 0.72 ppm) in the axial conformation of 14. This correlation is corroborated by the complementary but opposite shift differences, which are observed in the ¹³C-NMR spectra, the so-called γ -effect [11]. When the MeO group lies close to C(2) or C(6), shielding occurs. In the (*E*)-isomer 13, the resonance of C(6) appears upfield (Δ 4.43 ppm) from that of the (*Z*)-isomer 14 exhibits its C(2) resonance upfield (Δ 4.99 ppm) relative to that of the (*E*)-isomer 13.

Having established the geometric configurations of 13 and 14, they were converted by standard procedures into the corresponding 3'-oxobutyl, 3'-oxo-3'-phenylpropyl, and 3'-oxopropyl derivatives of the (E)- (15, 17, and 19) and (Z)-series (16, 18, and 20;Scheme 2).

Next, oxygenative cyclization was accomplished as before in two steps. First, the photo-oxygenation of 16 was assayed at low temperatures in different solvents to find the best conditions. The resulting solution was then treated with *Amberlyst-15 (Table 2)*. In each case, just three products (21–23) were isolated as colorless crystalline solids by chromatography (*Scheme 3*). Although elemental analysis indicated that they had all incorporated one molecule of O_2 , their structures could not be determined by NMR spectroscopy. Fortunately, recourse to X-ray analysis revealed that two were a pair of

Solvents		Catalyst	Products		
A ^a)	<i>B</i> ^b)		21	22	23
MeOH	CH ₂ Cl ₂	Amberlyst-15	10	10	17
MeCN ^d)	MeCN	Amberlyst-15	28	10	36
CH ₂ Cl ₂	CH ₂ Cl ₂	Amberlyst-15	48	19	30
CH ₂ Cl ₂	CH ₂ Cl ₂	AlCl ₃	39	17	0
CH ₂ Cl ₂	CH ₂ Cl ₂	Me ₃ SiOTf	16.6	10	0

Table 2. Oxygenated Cyclic Products (21–23) Obtained by Photo-oxygenation*) of (Z)-4-[2-(Methoxymethylidene)cyclohexyl]butan-2-one (16) and Isomerization^b) of the Intermediate 1,2-Dioxetane (24)^c)

^a) Photo-oxygenation was carried out in solvent A at -78° , except where noted. ^b) Catalyzed isomerization was carried out in solvent B at 0°. ^c) For further details, consult *Exper. Part.* ^d) Photo-oxygenation at -30° .



'endo'- and 'exo'-methoxy-1,2,4-trioxanes (21 and 22; Fig. 1), and that the third was a formyl peroxide (23; Fig. 2). The trioxane ring in the 'endo'- and 'exo'-epimers 21 and 22 exists in a twisted-boat conformation just like that observed in artemisinin (1) [12]. Furthermore, ring D, the dioxepane portion, adopts a twist-chair conformation which has a pseudo- C_2 axis passing through the O(1) atom [13]. In the bicyclic peroxide 23, the dioxepane ring still keeps the twist-chair conformation with the pseudo- C_2 axis cutting through the C(1) atom, and has the particularity of bearing *trans*-disposed CHO and MeOH substituents. The yield and proportions of the trioxanes and peroxide 21-23 depend crucially on the solvent and the catalysts used (*Table 2*). In MeOH, apart from 21-23, which were formed in minor amounts, a complex mixture of non-identifiable polar products was formed. On passing to the non-polar solvents MeCN and CH₂Cl₂, a net improvement occurred giving 21-23 in high yield. The use of AlCl₃ was superior to Me₃SiOTf as catalyst. Both had the advantage of not giving the undesired peroxide 23.

On following the photo-oxygenation of 16 by TLC, it was noticed that the 'endo'epimer 21 formed first and that the 'exo'-epimer 22 and peroxide 23 appeared later. This behavior suggests that 22 and 23 arise from 21, which, in fact, they do. In an independent experiment, 21 alone in CH_2Cl_2 was treated with *Amberlyst-15* at -10° for 5 h. It was transformed completely into the 'exo'-trioxane 22 and the peroxide 23.

When the photo-oxygenation of 16 was repeated in CH_2Cl_2 or $CDCl_3$ at -78° , but in the absence of acid catalyst, a single product was formed which appeared from its NMR spectrum to be 1,2-dioxetane 24 (*Scheme 4*). On warming the solution to room temperature, 2-(3'-oxobutyl)cyclohexanone (25), the scission product of 24, was obtained. This latter finding provides a rationale for the formation of trioxane 21. The structure postulated for 24 arises by attack of $^{1}O_2$ on the least hindered side of the double bond of 16



Fig. 1. Perspective drawings of the crystal structures of the 'endo'- and 'exo'-methoxy-1,2,4-trioxanes 21 and 22, respectively, with atomic numbering (arbitrary). Ellipsoids are represented with 40% probability.



Fig. 2. Perspective drawings of the crystal structure of peroxide 23 with atomic numbering (arbitrary). Ellipsoids are represented with 40% probability.

which, for purposes of discussion, is shown as the (R)-enantiomer (Scheme 4). Conformational inversion places the side chain close to the dioxetane grouping, the future reactive site $(24 \rightarrow 24')$. Protonation and opening of the dioxetane ring in 24' creates 26, the methylideneoxonium moiety of which is immediately attacked on its *si* face by the pendent carbonyl function. The latter is simultaneously attacked by the newly formed hydroperoxy group thereby completing the bicyclic closure and producing the kinetic product, the 'endo'-epimer 21. Subsequent protonation of 21 affords the tricyclic cation 27 and a molecule of MeOH which have the chance of recombining on the *re* or *si* faces of the methylideneoxonium moiety, thereby accounting for the equilibration of the 'endo'and 'exo'-epimers 21 and 22 (Scheme 4). However, a third avenue is possible. Apo-attack cleaves the trioxane ring with Walden inversion to place the entering MeO substituent uniquely *trans* to the newly formed aldehyde group ($27 \rightarrow 23$). Interestingly, when nonprotic catalysts were used, the peroxide 23 was not observed (*Table 2*). This result could mean that the *Lewis* acid tightly binds to the MeO group and the site from whence it came so preventing the more complicated maneuver required for ring cleavage.



Me

٨e

re

The same sequence of photo-oxygenation and acid catalysis was carried out with the (E)-isomer 15. By choosing nonpolar solvents, the yields of products were better than 65%, but in addition to 21–23, cyclohexenecarbaldehyde 30 appeared as the major product (*Scheme 5*). The provenance of 30 is a consequence of the preferred equatorial



2780

conformation of 15 which, unlike 16, has axial allylic H-atoms available for a competing ene reaction. Attack by ${}^{1}O_{2}$ on the least hindered face, therefore, follows two courses. Cycloaddition as before is expected to give dioxetane 28. Subsequent acid-catalyzed isomerization implicating the keto side-chain would logically give the '*exo*'-methoxy epimer 22 as the kinetic product, but this point was not verified experimentally. Thereafter, the '*endo*'-methoxy epimer 21 and peroxide 23 would arise by acid-catalyzed equilibration. The incoming molecule of ${}^{1}O_{2}$ as it approaches the double bond of 15 also experiences the '*syn*'-effect exerted by the MeO group [14]. The ene reaction supervenes and the axial H-atom at the least substituted adjacent center is abstracted to give hydroperoxide 29 and eventually 30 by hydrolysis during the workup.

Submission of the phenyl-substituted (Z)- and (E)-enol ethers 18 and 17 to photooxygenation and Amberlyst-catalyzed rearrangement also resulted in capture of an O_2 molecule, but less efficiently (Scheme 6). The (Z)-isomer 18 gave the 'endo'- and 'exo'methoxy-1,2,4-trioxanes 32 and 33, accompanied by the peroxide 34. Evidence for the intermediate 1,2-dioxetane 31 was also secured. Only the 'endo'-epimer 33 and its cleavage product 34, together with aldehyde 35 arising from the ene reaction, were obtained from the (E)-epimer 17.



Lastly, the 3'-oxopropyl-substituted (Z)-enol derivative 20 was photo-oxygenated directly without prior purification in view of its instability (*Scheme 7*). Once again, evidence for the 1,2-dioxetane 36 was obtained. Acid catalysis converted it to the 'endo'- and 'exo'-methoxy epimers 37 and 38 in 22 and 30% yield, respectively. No trace of the corresponding bicyclic peroxide was detected. Similar treatment of the (E)-isomer 19 was disappointing: no trioxanes were isolated, and only extensive decomposition of the reaction mixture was observed.

In vitro Antimalarial Activity. – Samples of the synthetic tricyclic 1,2,4-trioxanes and appropriate reference compounds were tested *in vitro* against *Plasmodium falciparum* clones and strains by using the method developed by *Desjardins* and coworkers, and subsequently modified by *Milhous* and other authors [15] [16]. Unfortunately, the ACD derivatives **37** and **38** were not sufficiently stable to be tested.

The effectiveness of the sample in inhibiting nucleic-acid synthesis as measured by the incorporation of tritiated hypoxanthine in the clones and strains was determined. Inhibitory concentrations (IC) are expressed in nanogram/ml. The Indochina W2 clone is resistant to chloroquine, quinine, pyrimethamine, and sulfadoxine but susceptible to mefloquine, whereas the Sierra Leone D6 clone is susceptible to chloroquine, quinine, pyrimethamine, and sulfadoxine. The Kanchanaburi K1 strain is resistant to chloroquine and pyrimethamine [17] and the CDCI Honduras strain is sensitive to chloroquine and resistant to pyrimethamine [18].

	W2 clone		D6 clone		
	<i>IC</i> ₅₀ [ng/ml]	<i>IC</i> ₉₀ [ng/ml]	<i>IC</i> ₅₀ [ng/ml]	<i>IC</i> ₉₀ [ng/ml]	
1	1.1	-	2.2	***	
10	6.2	25.8	28.7	59.8	
11	2.0	3.3	2.3	30.4	
21	1.8	3.9	16.5	11.5	
22	9.7	16.9	754	875	
23	1837	8132	2608	7547	
Quinine	9.6	-	2.1	_	
Chloroquine	23.4	-	0.9	_	
Mefloquine	0.8	-	4.6	-	
Pyrimethamine	5.5	-	0.01	_	
Sulfadoxine	2060		35		

Table 3. In vitro Antimalarial Activity of Some Artemisinin Analogues Against P. falciparum Clones

Table 4. In vitro Antimalarial Activity of Some Artemisinin Analogues Against P. falciparum Strains

	K1-strain		Honduras strain		
	<i>IC</i> ₅₀ [ng/ml]	<i>IC</i> ₉₀ [ng/ml]	<i>IC</i> ₅₀ [ng/ml]	<i>IC</i> ₉₀ [ng/ml]	
1	1.6 (1.3-1.9)	4.0 (3.3-4.9)	1.1 (0.6-1.8)	3.4 (2.0-5.6)	
21	3.6 (2.7-4.7)	22.0 (16.6-29.3)	6.4 (4.6-8.8)	51.8 (38.0-71.7)	
22	40.5 (38.6-42.6)	103.2 (98.3-108.4)	49.5 (35.9-68.1)	236.3 (171.6-325.8)	
32	4.0 (2.0-8.0)	18.2 (9.0-36.5)	9.2 (7.2-11.7)	32.3 (25.4-41.1)	
33	9.6 (6.8-13.4)	34.0 (24.2-47.6)	10.5 (7.9-14.0)	39.6 (29.8-52.7)	
Chloroquine	87.1 (69.6-109.5)	221.4 (176.4-277.9)	5.7 (5.0-6.5)	10.2 (8.9–11.8)	

The IC_{50} and IC_{90} values determined in these assays (*Tables 3* and 4) provide a measure of the efficacy of a particular compound with respect to a panel of conventional quinoline- and sulfonamide-based drugs.

2782

Discussion. – It is no surprise that the synthetic trioxanes which display high activities are those (10 and 11) possessing a part of the molecular architecture of artemisinin (1) (*Table 3*). Their extraordinarily small IC_{50} and IC_{90} values confirm that ring D and a lactone function are not essential for activity. The difference in activity between 10 and 11, although not great, appears to be significant. Clearly, the Me₂C grouping in 10 serves as a substitute for the ethane bridge in ring D of 1. However, its replacement by the spirocyclopentane group is even more effective (see 11) and probably springs from the conformational constraint or compression that it exerts on the attached trioxane ring causing it to buckle into a boat-like form.

Remarkably high activities are also exhibited by the pair of epimers 21 and 22. The very small IC_{50} and IC_{90} values confirm again that the lactone ring is not part of the pharmacophore and that even ring B is not necessary for artemisinin-like activity. It is worth noting that although the tetrahydropyran ring, *e.g.* ring B in 10 and 11, can be dispensed with, a fourth O-atom is still required. In 21 and 22, the MeO substituent fulfills this requirement. Paradoxically, the '*exo*'-methoxy substituent in 22, which mimics the orientation of the O-atom in the B ring of artemisinin (1) and the *seco*artemisinins 10 and 11, confers less activity than its '*endo*'-epimer 21.

The large IC_{50} and IC_{90} values for 23 demonstrate that an intact trioxane ring is a *sine qua non* and that the cyclic peroxide function alone is insufficient for antimalarial activity. This result also suggests that the aforementioned difference in efficacy between the 'endo'- and 'exo'-epimers 21 and 22, respectively, might be due to the decomposition of 22 to 23 by traces of acid.

The synthetic 1,2,4-trioxanes follow the same trend as that of artemisinin in being more effective against the chloroquine-resistant W2 clone. The most potent trioxanes are 11 and 21 in that they have the smallest IC_{50} and IC_{90} values. In addition, the activities of 21 and 22 compare well with that of the chiral natural product, artemisinin, especially when it is remembered that they are not only fully synthetic but racemic mixtures.

The relative activities of 21 and 22 against the *P. falciparum* clones were confirmed in the K1 and Honduras strains (*Table 4*). Replacement of the bridgehead Me group by the Ph substituent improved antimalarial action. Comparison of the 'endo'-epimers 21 and 32 shows that they have about the same activity against the K1 and Honduras strains. More importantly, the difference between the Ph-substituted 'endo'- and 'exo'-methoxy epimers (32 vs. 33) is in the same sense as that seen for the Me analogues (21 and 22). Moreover, the Ph-substituted 'exo'-methoxy trioxane 33 is superior in activity to its Me homologue 22. Once again, the trioxanes are more effective against the chloroquine-resistant than the sensitive strain.

In general, all new trioxanes performed better than the traditional N-containing drugs, with the exception of mefloquine. Similar results as those reported for **21** and **32** here and earlier [19] were also obtained with related hydroxyethyl derivatives [20]. However, in these experiments, only the '*endo*'-methoxy epimers were isolated and tested. Equilibration of the '*endo*'- to the '*exo*'-epimers and eventually to the cleaved peroxide products was not reported. Furthermore, two methylated lactone analogues of **10** were reported to have comparable *in vitro* activity [21].

The present findings indicate that possible pharmacophores for high artemisinin-like activity may be represented by the hypothetical doubly spirocyclic trioxane **39** and the *cis*-fused bicyclic entity **40**. It is also to be noted that a Ph substituent, although not part



of the pharmacophore, but attached to the trioxane ring, is beneficial for activity. Apart from these structural requirements, chemical stability, particularly towards acid, is also a prerequisite for a candidate antimalarial drug. The homochiral ABC compounds 10 and 11 are certainly stable enough, but are relatively inaccessible as their syntheses entail many steps. On the other hand, the easily prepared racemic ACD derivatives 21, 22, 32, and 33 are potentially disadvantaged by their equilibration to less active isomers when acids are present. Studies such as those described here have played an important part in the design of simple *cis*-fused bicyclic 1,2,4-trioxanes which are stable and potent antimalarial drug candidates [22].

We gratefully acknowledge the UNDP/World Bank/WHO Special Programme for Research and Training in Tropical Diseases for partial support of this research. We also thank Messrs J. P. Saulnier and A. Pinto for the NMR measurements, and Mrs C. Clément for the MS measurements.

Experimental Part

General. Solvents were purchased from Fluka AG and Aldrich AG; when necessary, they were dried and purified before use. TLC: plates coated with silica gel 60 (F_{254} Merck). Flash column chromatography (FC): silica gel 60 (230–400 mesh, Merck); CC = column chromatography. M.p. (uncorrected): Reichert hot-plate instrument. IR Spectra: Perkin-Elmer-681 and FT-M-Polaris spectrometers; in CHCl₃. NMR Spectra: Varian-XL-200 and Bruker-WH-360 instruments; in CDCl₃; chemical shifts (δ) in ppm with reference to Me₄Si; signal intensities are normalized to 1 H; coupling constants J in Hz. MS: Finnigan-4000 and VG-70-70E spectrometers. Elemental analyses were performed by Dr. H.J. Eder, Service de Microchimie, Département de Chimie Pharmaceutique, University of Geneva.

Procedure for photo-oxygenation: A soln. of the reactant was cooled to -78° with a dry-ice/acetone bath or a *Huber-TC-50* cryostat for other temperatures, while O₂ was passed. Irradiation was effected with a tungsten filament lamp (*Sylbania FFX*), cooled by a jet of compressed air, and screened with a *Spindler* and *Hoyer GG 475* filter to cut off UV light.

1. (E)- and (Z)-3-[2-(Methoxymethylidene)cyclohexyl]propanenitrile (13 and 14, resp.). A soln. of (methoxymethyl)diphenylphosphine oxide (12.3 g, 50 mmol) [8] in THF (300 ml) was stirred at 0° for 20 min with lithium diisopropylamide (LDA) prepared from (i-Pr)₂NH (5.05 g, 50 mmol) and 1.6M BuLi/hexane (31.25 ml) in THF (60 ml). The soln. was cooled to -70° , and 3-(2-oxocyclohexyl)propanenitrile (12; 7.55 g, 50 mmol) in THF (50 ml) was added dropwise [23]. The resulting mixture was allowed to warm to 20° and then stirred further for 24 h. Next, hexane (100 ml) was added. Removal of the precipitated diphenylphosphinite by filtration over *Celite* gave a soln. which was dried (MgSO₄) and evaporated. The oily residue was purified by CC (silica gel, 20% AcOEt/pentane): 13 (R_f 0.37; 2.68 g, 30%) and 14 (R_f 0.42; 2.7 g, 30%). Colorless oils.

13: IR: 2938*s*, 2830*m*, 2820*w*, 2245*m*, 1125*s*. ¹H-NMR: 1.0–1.9 (*m*, 9 H); 2.0 (*m*, 1 H); 2.1 (*m*, 2 H); 2.3 (*dt*, 1 H); 3.48 (*s*, 3 H); 5.75 (*s*, 1 H). ¹³C-NMR: 140.37; 119.91; 117.47; 59.38; 37.54; 32.74; 27.01; 26.91; 22.38; 21.88; 15.29. Anal. calc. for C₁₁H₁₇NO: C 73.69, H 9.56, N 7.82; found: C 71.89, H 9.21, N 7.52.

14: IR: 2940s, 2860m, 2840w, 2245m, 1680m, 1125s. ¹H-NMR: 1.2–2.3 (*m*, 12 H); 2.9 (*m*, 1 H); 3.52 (*s*, 3 H); 5.86 (*s*, 1 H). ¹³C-NMR: 141.37; 120.53; 116.9; 59.22; 32.55; 31.31; 28.02; 27.57; 26.31; 21.70; 15.21. Anal. calc. for $C_{11}H_{17}NO$: C 73.69, H 9.56, N 7.82; found: C 71.54, H 9.18, N 7.72.

 δ (H) and δ (C) of H–C(2) and H–C(6) of 13 and 14 were assigned unambiguously by means of DEPT and hetero-COSY experiments (see *Table 1*).

2. (E)- and (Z)-4-[2-(Methoxymethylidene) cyclohexyl]butan-2-one (15 and 16, resp.). A soln. of 13 (200 mg, 1.11 mmol) in dry Et₂O (20 ml) was stirred at 0° under N₂ to which 1.6M MeLi/Et₂O (1.4 ml) was added dropwise. The mixture was stirred for further 6 h. Thereafter, a sat. aq. soln. of NH₄Cl (10 ml) was added dropwise with stirring. Next, the org. layer was separated, the aq. phase extracted with Et₂O (2 × 10 ml), and the combined org. layer washed (H₂O), dried (MgSO₄), and evaporated. CC (SiO₂, 2% AcOEt/pentane) gave 15 (200 mg, 91%). Yellow oil. R_f 0.35. IR: 3000w, 2936s, 2860m, 1720s, 1680m, 1125s, 1095w. ¹H-NMR: 1.2–2.0 (m, 10 H); 2.1 (s, 3 H); 2.35 (m, 3 H); 3.45 (s, 3 H); 5.72 (s, 1 H). Anal. calc. for C₁₂H₂₀O₂: C 73.41, H 10.28; found: C 73.19, H 10.30.

A soln. of **14** was treated with MeI in exactly the same way as **13**: **16** (198 mg, 90%). Oil. R_f (0.32. IR: 3000w, 2940s, 2860m, 2840w, 1710s, 1680m, 1125s, 1095m. ¹H-NMR: 1.2–2.0 (m, 10 H); 2.12 (s, 3 H); 2.4 (m, 2 H); 2.8 (m, 1 H); 3.5 (s, 3 H); 5.8 (s, 1 H). Anal. calc. for C₁₂H₂₀O₂: C 73.41, H 10.28; found: C 73.21, H 10.28.

3. (E)- and (Z)-3-[2-(Methoxymethylidene)cyclohexyl]-3-phenylpropan-1-one (17 and 18). The enol ethers 13 and 14 (400 mg, 2.22 mmol) were dissolved separately in Et₂O (30 ml) at 0°, and each was treated as described below. PhLi (2.2 ml, 2.0m in benzene/Et₂O; 2 equiv.) was the added dropwise under N₂, the mixture stirred for 3 h at 0°, and sat. aq. NH₄Cl soln. (20 ml) added. The aq. layer was then extracted with Et₂O, the Et₂O layer washed, dried (MgSO₄), and evaporated, and the residue submitted to FC (SiO₂, CH₂Cl₂) [24]: 17 (450 mg, 78%: R_f 0.39) and 18 (500 mg, 86%: R_f 0.41).

17: IR: 3000w, 2930s, 2860m, 1680s, 1600w, 1120s. ¹H-NMR: 1.3–2.1 (m, 10 H); 2.35 (m, 1 H); 3.0 (t, ³J = 7, 2 H); 3.56 (s, 3 H); 5.8 (s, 1 H); 7.5 (m, 3 H); 8.0 (m, 2 H). Anal. calc. for C₁₇H₂₂O₂: C 79.02, H 8.59; found: C 78.85, H 8.42.

18: IR : 3000w, 2940s, 2860m, 2840w, 1680s, 1600w, 1125s. ¹H-NMR : 1.0–2.1 (m, 10 H); 2.95 (m, 1 H); 3.0 (t, ³J = 7, 2 H); 3.48 (s, 3 H); 5.84 (d, ⁴J = 1.5, 1 H); 7.5 (m, 3 H); 8.0 (m, 2 H). Anal. calc. for C₁₇H₂₂O₂: C 79.02, H 8.59; found: C 78.59, H 8.77.

4. (E)- and (Z)-3-[2-(Methoxymethylidene)cyclohexyl]propanal (19 and 20, resp.). To solns. of 13 and 14 (100 mg, 0.56 mmol), dissolved separately in THF (10 ml) at -78° , was added 2M AlH(i-Bu)₂/hexane (0.8 ml, 1.5 equiv.). The resulting soln. was allowed first to warm to 20° and then stirred for 12 h. Next, a sat. aq. soln. of NH₄Cl was added, the aq. phase extracted with Et₂O, and the combined Et₂O extract washed, dried (MgSO₄), and evaporated. The ¹H-NMR and IR spectra of the residue revealed the formation of 19 (80 mg, 79%) and 20 (85 mg, 85%).

19: $R_{\rm f}$ 0.67 (10% AcOEt/CH₂Cl₂). IR: 2920s, 2860m, 1725s, 1680m, 1125s, 1100w. ¹H-NMR: 1.0–2.2 (m, 13 H); 3.52 (s, 3 H); 5.78 (br. s, 1 H); 9.82 (t, ³J = 2, 1 H).

20: $R_{\rm f}$ 0.65 (10% AcOEt/CH₂Cl₂). IR: 2925*s*, 2860*m*, 1720*s*, 1680*m*, 1120*s*, 1095*w*. ¹H-NMR: 1.0–2.1 (*m*, 12 H); 2.9 (*m*, 1 H); 3.5 (*s*, 3 H); 5.8 (br. *s*, 1 H); 9.8 (*t*, ³*J* = 2, 1 H).

5. Photo-oxygenation of 16. 5.1. In MeOH. A soln. of 16 (100 mg, 0.51 mmol) and methylene blue (MB; 5 mg) in MeOH (10 ml) was photo-oxygenated at -78° for 2 h. TLC (5% AcOEt/CH₂Cl₂): 1 product spot. $R_{\rm f}$ 0.32. The MeOH was evaporated in vacuo at -30° and the residue taken up in CH₂Cl₂ at -50° . Amberlyst-15 (200 mg) was added and the mixture stirred for 10 h at 0°. Removal of the catalyst by filtration and separation by CC (SiO₂, CH₂Cl₂) afforded 21 (12 mg, 10%). 22 (12 mg, 10%), and 23 (20 mg, 17%) as crystalline colorless solids.

5.2. In MeCN. A soln. of **16** (100 mg, 0.51 mmol) and MB (5 mg) in MeCN (10 ml) was photo-oxygenated at -30° for 1 h. Amberlyst-15 (100 mg) was added to the soln. which was then stirred for 5 h at 0°. The previous workup was followed to give **21** (32 mg, 28%), **22** (12 mg, 10.5%), and **23** (42 mg, 36%).

5.3. In CH_2Cl_2 . A soln. of 16 (100 mg, 0.51 mmol) and MB (5 mg) in CH_2Cl_2 (10 ml) was photo-oxygenated at -78° for 1 h, after which time *Amberlyst-15* (100 mg) was added. The mixture was stirred at 0° for 10 h and submitted to the usual workup to give 21 (56 mg, 48%), 22 (22 mg, 19%), and 23 (35 mg, 30%).

5.4. Without Acid Catalyst. The photo-oxygenation as described in 5.3 was repeated with **16** (20 mg, 0.1 mmol) in CH₂Cl₂ (10 ml). After stirring the mixture for 1 h, a portion was examined by TLC (25% AcOEt/CH₂Cl₂): 1 product spot, R_f 0.5. The solvent was then evaporated to give (2RS)-2-(3'-oxobutyl)cyclohexanone (**25**; 15.5 mg, 90%) as colorless liquid [25].

5.5. In CDCl₃. The same procedure was followed as in 5.3, but in CDCl₃ as solvent. The NMR spectrum of the mixture at -35° gave ¹H-NMR signals characteristic of dioxetane 24.

5.6. Catalysis with AlCl₃ and Me_3SiOTf . Photo-oxygenation of **16** was performed as in 5.3. AlCl₃ (50 mg) was added to the soln. at 0°, followed by neutralization with a few drops of Et₃N. Workup gave **21** (48 mg, 39%) and **22** (20 mg, 17%). The same procedure but using Me₃SiOTf (0.1 ml) as the *Lewis* acid also gave **21** (20 mg, 17%) and **22** (12 mg, 10%).

*4-[(3*RS,*4*RS,*5*SR)-*3-Methoxy-1,2-dioxaspiro[3.5]nonan-5-yl]butan-2-one* (**24**): ¹H-NMR: 1.0-2.6 (*m*, 13 H); 2.2 (br. *s*, 3 H); 3.41 (*s*, 3 H); 5.25 (br. *s*, 1 H).

(1 RS, 6 SR, 9 SR, 12 RS)-12-Methoxy-9-methyl-10,11,13-trioxatricyclo[7.2.2.0^{1,6}]tridecane (21): M.p. 74–75° (from pentane). $R_{\rm f}$ 0.5. IR: 3000w, 2940s, 2860m, 1450m, 1140s, 1120s, 1030s. ¹H-NMR: 1.25 (m, 5 H); 1.65–1.85 (m, 8 H); 2.05 (ddd, ²J = 15, ³J = 4, 3, 1 H); 2.3 (ddd, ²J = 15, ³J = 14, 4, 2 H); 3.53 (s, 3 H); 4.93 (s, 1 H). MS: no M^+ , 196 (0.8), 168 (3.4), 150 (2.0), 138 (9), 125 (100), 111 (20), 98 (22), 81 (23), 55 (95). Anal. calc. for C₁₂H₂₀O₄: C 63.12, H 8.84; found: C 62.99, H 8.93.

 $(1 \text{ RS}, 6 \text{ SR}, 9 \text{ SR}, 12 \text{ SR}) - 12 - Methoxy - 9 - methyl - 10, 11, 13 - trioxatricyclo[7.2.2.0^{1.6}] tridecane (22): M.p. 63° (from hexane). R_f 0.19. IR : 3000w, 2945s, 2860m, 1450m, 1145s, 1120s, 1025s. ¹H-NMR: 1.2 (m, 5 H); 1.4 (s, 3 H); 1.7 (m, 4 H); 1.85 (m, 1 H); 2.0 (m, 1 H); 2.4 (m, 2 H); 3.5 (s, 3 H); 4.95 (s, 1 H). MS: no <math>M^+$, 196 (10), 168 (17), 138 (50), 125 (98), 111 (42), 98 (42), 55 (100). Anal. calc. for $C_{12}H_{20}O_4$: C 63.12, H 8.84; found: C 63.38, H 8.99.

(3 RS, 5a SR, 9a RS) - 3, 4, 5, 5a, 6, 7, 8, 9- Octahydro-3-methoxy-3-methyl-9a H-1,2-benzodioxepine-9a-carbalde-hyde (23): M.p. 80° (from hexane). $R_f 0.3$. IR: 3000w, 2940s, 2860m, 1740s, 1450m. ¹H-NMR: 1.2 (s, 3 H); 1.21–2.1 (m, 13 H); 3.35 (s, 3 H); 9.55 (d, 1 H). Anal. calc. for $C_{12}H_{20}O_4$: C 63.12, H 8.84; found: C 63.07, H 8.87.

Crystallographic Data for 21–23. Cell parameters and diffracted intensities were measured at r.t. on a Philips PW1100 diffractometer with graphite-monochromated Mo[$K\alpha$] radiation ($\lambda = 0.71069$ Å). Data were corrected for Lorentz and polarization effects, but not for absorption. During the data collections, the diffracted intensities decreased by ca. 2, 17, and 37% for 21, 22, and 23, resp. All intensities were corrected for these drifts. The structures were solved by direct methods using MULTAN 80 [26], all other calculations used XRAY-76 [27] system and ORTEP [28] programs. All coordinates of the H-atoms were calculated. A summary of crystal data, intensity measurement, and structure refinement is givewn in Table 5 and selected geometrical parameters are reported in Table 6. Crystallographic data were deposited with the Cambridge Crystallographic Data Center, University Chemical Laboratory, 12 Union Road, Cambridge CB2 1EW, England.

	21	22	23
Formula	$C_{12}H_{20}O_4$	$C_{12}H_{20}O_4$	C ₁₂ H ₂₀ O ₄
Mol. wt.	228.3	228.3	228.3
Crystal system	triclinic	monoclinic	triclinic
Space group	$P\overline{1}$	$P2_1/c$	P 1
a [Å]	6.9908(8)	16.807(3)	6.462(3)
<i>b</i> [Å]	7.9224(6)	5.984(1)	10.393(1)
c [Å]	12.266(2)	13.092(2)	10.652(5)
α [°]	75.02(2)	90	63.15(4)
β[°]	84.71(2)	111.56(1)	81.18(4)
γ [°]	64.99(3)	90	75.19(2)
$V[Å^3]$	594.6(2)	1224.6(4)	616.5(5)
Ż	2	4	2
F(000)	248	496	248
$D_{\rm c} \left[{\rm g} \cdot {\rm cm}^{-3} \right]$	1.28	1.24	1.23
$\mu(MoK_{\gamma}) [mm^{-1}]$	0.088	0.086	0.085
$((\sin\theta)/\lambda)_{\max}$ [Å ⁻¹]	0.53	0.51	0.51
No. measured reflections	1445	1528	1328
No. observed reflections	1231	759	874
Criterion for observed	$ F_{o} > 4\sigma(F_{o})$	$ F_{\rm o} > 4\sigma(F_{\rm o})$	$ F_{\rm o} > 4\sigma(F_{\rm o})$
Refinement (on F)	full-matrix	full-matrix	full-matrix
No. parameters	145	145	145
Weighting scheme	$\omega = 1$	$\omega = 1$	$\omega = 1$
Max. and average Δ/σ	0.0016, 0.0003	0.011, 0.02	0.0006, 0.0001
Max. and min. $\Delta \rho [e \cdot Å^{-3}]$	0.17, -0.31	0.35, -0.45	0.27, -0.35
S	7.8	5.7	4.6
$R(=\omega R)$	0.049	0.080	0.079

Table 5. Summary of Crystal Data, Intensity Measurement, and Structure Refinement for 21-23

	21	22	23		21	22	23
C(1)-O(1)	1.417(5)	1.45(2)	1.44(1)	O(3)-C(1)-O(1)-O(2)	-71.5(3)	-73.4(8)	
O(1)-O(2)	1.473(4)	1.47(1)	1.467(7)	C(1) - O(1) - O(2) - C(3)	39.9(3)	43(1)	108.6(8)
O(2)-C(3)	1.473(5)	1.48(1)	1.45(1)	O(1) - O(2) - C(3) - C(2)	23.0(3)	20(1)	44.2(8)
C(3) - C(2)	1.525(6)	1.50(1)	1.52(1)	O(2) - C(3) - C(2) - O(3)	-61.5(3)	-57(1)	-138.1(9)
C(2)-O(3)	1.427(4)	1.43 (1)	1.18(2)	C(3)-C(2)-O(3)-C(1)	32.8(4)	27(1)	-
O(3)-C(1)	1.424(5)	1.47(1)	-	C(2) - O(3) - C(1) - O(1)	32.1(3)	35(1)	-
O(4) - C(1)	-	-	1.41(1)	O(3)-C(2)-O(4)-C(12)	71.1(3)	-61(1)	
O(4)-C(2)	1.391(5)	1.40(1)	-	C(3)-C(2)-O(4)-C(12)	-166.7(3)	173.0(9)	-
				O(1)-O(2)-C(3)-C(8)	-102.6(3)	-103.3(9)	-82.9(7)
O(3)-C(1)-O(1)	107.7(2)	106(1)	-	C(3)-C(8)-C(9)-C(10)	-41.9(5)	-40(1)	-72(1)
C(1)-O(1)-O(2)	110.1(2)	109.8(7)	111.5(6)	C(8)-C(9)-C(10)-C(1)	58.6(5)	57(1)	85(1)
O(1) - O(2) - C(3)	112.5(3)	111.6(7)	105.3(6)	C(9)-C(10)-C(1)-O(1)	-95.0(4)	-95(1)	-34(1)
O(2)-C(3)-C(2)	106.2(3)	109.7(7)	106.6(7)	C(10)-C(1)-O(1)-O(2)	50.8(3)	49(1)	-51(1)
C(3) - C(2) - O(3)	112.5(2)	113.7(8)	128(1)				
C(2)-O(3)-C(1)	113.3(3)	113.7(9)	-				
C(2)-O(4)-C(12)	113.0(3)	113.9(9)	-				

Table 6. Selected Bond Lengths [Å], Bond Angles [°] and Torsional Angles [°] for 21-23

6. Photo-oxygenation of 15. 6.1. In MeCN. A soln. of 15 (90 mg, 0.46 mol) and MB (5 mg) in MeCN (10 ml) was photo-oxygenated at -50° for 1 h. Amberlyst-15 (50 mg) was then added and the mixture stirred at -20° for 5 h. On workup, 21 (12 mg, 11%), 22 (9 mg, 9%), and 23 (12 mg, 11%) were obtained, together with 6-(3'-oxobutyl)cyclohex-1-ene-1-carbaldehyde (30) as a colorless oil.

6.2. In CH_2Cl_2 . The procedure in 6.1 was followed, except that CH_2Cl_2 was used as solvent and the temp. kept at -78° . On workup, **21** (18 mg, 17%), **22** (18 mg, 17%), **23** (15 mg, 14%), and **30** (27 mg, 30%) were obtained.

30: IR: 2940s, 2860m, 2320m, 1715s, 1685s. ¹H-NMR: 1.4–1.9 (m, 8 H); 2.15 (s, 3 H); 2.2–2.5 (m, 3 H); 6.8 (t, ${}^{3}J = 4, 1$ H); 9.4 (s, 1 H).

7. Photo-oxygenation of 18. A soln. of 18 (100 mg, 0.39 mmol) and MB (10 mg) in CH₂Cl₂ (20 ml) was photo-oxygenated at -78° for 45 min. TLC (10% AcOEt/CH₂Cl₂): presence of the corresponding 1,2-dioxetane 31, $R_{\rm f}$ 0.06. Amberlyst-15 (80 mg) was then added and the mixture stirred at 0° for 6 h. After workup, the crude product was subjected to FC [24] (SiO₂, 10% AcOEt/CH₂Cl₂): 32 (10 mg, 9%), 33 (18 mg, 16%), and 34 (14 mg, 12.5%) as a colorless oil, solid, and oil, resp.

 $(1 \text{ RS}, 6 \text{ SR}, 9 \text{ RS}, 12 \text{ RS}) - 12 - Methoxy-9-phenyl-10, 11, 13-trioxatricyclo[7.2.2.0^{1.6}] tridecane (32): R_f 0.7. IR: 3100w, 2940s, 2860m, 1450m, 1100s, 1010s. ¹H-NMR: 1.25-2.2 (m, 11 H); 2.35 (ddd, ²J = 10, ³J = 5, 3.5, 1 H); 2.8 (ddd, ²J = 15, ³J = 13, 4, 1 H); 3.68 (s, 3 H); 5.15 (d, ⁴J = 1.5, 1 H); 7.4 (m, 3 H); 7.6 (m, 2 H). Anal. calc. for C₁₇H₂₂O₄: C 70.31, H 7.64; found: C 70.18, H 7.63.$

(1 RS, 6 SR, 9 RS, 12 SR) - 12-Methoxy-9-phenyl-10,11,13-trioxatricyclo[7.2.2.0^{1.6}]tridecane (**33**): M.p. 82°. R_f 0.5. IR: 3150w, 2935s, 2860m, 1450m, 1100s, 1015s. ¹H-NMR: 1.2–2.2 (m, 10 H); 2.3 (m, 1 H); 2.45 (m, 1 H); 2.9 (ddd, ²J = 15, ³J = 14, 4, 1 H); 3.65 (s, 3 H); 5.22 (s, 1 H); 7.4 (m, 3 H); 7.6 (m, 2 H). Anal. calc. for C₁₇H₂₂O₄: C 70.31, H 7.64; found: C 70.19, H 7.68.

(3 RS, 5a RS, 9a SR) - 3,4,5,5a,6,7,8,9 - Octahydro - 3 - methoxy - 3 - phenyl - 9a H - 1,2 - benzodioxepine - 9a - carbalde-hyde (34): R_f 0.61 (10% AcOEt/CH₂Cl₂). IR: 3030w, 2945s, 2860m, 1720s, 1280s, 1120m, 1100m, 1025s. ¹H-NMR: 1.2–2.2 (m, 11 H); 2.3 (ddd, ²J = 14, ³J = 7.5, 1.5, 1 H); 2.4 (ddd, ²J = 14, ³J = 11, 1.5, 1 H); 3.2 (s, 3 H); 7.34 (m, 4 H); 7.6 (m, 1 H). Anal. calc. for C₁₇H₂₂O₄: C 70.31, H 7.64; found: C 69.55, H 7.45.

8. Photo-oxygenation of 17. The procedure and quantities adopted for 17 were exactly the same as those described in *Exper.* 7. FC [24] (10% AcOEt/CH₂Cl₂) gave 33 (8 mg, 7%), 34 (8 mg, 7%), and 6-(3'-oxo-3'-phenyl)-cyclohex-1-ene-1-carbaldehyde (35; 33.8 mg, 36%).

35: Oil. $R_{\rm f}$ 0.25. IR: 3100w, 2940s, 2860m, 1705vs, 1685vs, 1580w, 1600w, 1450m. ¹H-NMR: 1.5–2.5 (m, 8 H); 2.7 (m, 1 H); 3.0 (m, 2 H); 6.8 (t, ³J = 3.5, 1 H); 7.5 (m, 3 H); 8.0 (m, 2 H); 9.4 (s, 1 H).

9. Photo-oxygenation of 20. A soln. of 20 (70 mg, 0.385 mmol) and MB (10 mg) in CH_2Cl_2 (20 ml) was photo-oxygenated at -78° for 1 h. TLC (10% AcOEt/CH₂Cl₂): 1,2-dioxetane 36 as the main product, R_f 0.6. Next,

Amberlyst-15 (15 mg) was added with stirring at -20° for 6 h (TLC: several products). Removal of the catalyst by filtration and submission of the filtrate to FC [24] (SiO₂, 3% AcOEt/CH₂Cl₂) furnished **37** (18 mg, 22%) and **38** (24 mg, 30%), together with other more polar, unidentified products.

 $(1 \text{ RS}, 6 \text{ SR}, 9 \text{ SR}, 12 \text{ RS}) - 12 - Methoxy - 10, 11, 13 - trioxatricyclo[7.2.2.0]^{1.6}$]tridecane (37): $R_{\rm f}$ 0.56. IR: 2940s, 2860w, 1095s, 1030m, 950m. ¹H-NMR: 1.1–2.1 (m, 11 H); 2.2 (m, 2 H); 3.5 (s, 3 H); 4.95 (s, 1 H); 5.65 (dd, ³J = 7.5, ⁴J = 1.5, 1 H). Anal. calc. for C₁₁H₁₈O₄: C 61.65, H 8.45; found: C 61.96, H 8.75.

 $(1 \text{ RS}, 6 \text{ SR}, 9 \text{ SR}, 12 \text{ SR}) - 12 - Methoxy - 10, 11, 13 - trioxatricyclo[7.2.2.0^{1.6}] tridecane ($ **38**): R_f 0.48. IR: 2935s, 2860w, 1100s, 1035m, 955m. ¹H-NMR: 1.1–2.5 (m, 11 H); 2.2 (m, 2 H); 3.55 (s, 3 H); 4.95 (s, 1 H); 5.6 (d, ³J = 8.0, 1 H). Anal. calc. for C₁₁H₁₈O₄: C 61.65, H 8.47; found: C 60.96, H 8.76.

REFERENCES

- 'Malaria, Obstacles and Opportunities', Eds. S.C. Oaks, Jr., V.S. Mitchell, G.W. Pearson, and C.C.J. Carpenter, National Academy Press, Washington D.C., 1991; 'Tropical Disease Research, Eighth Programme Report', Eds. J. Maurice and A. M. Pearce, World Health Organization, Geneva, 1987; W. Peters, Br. Med. Bull. 1982, 38, 187; W. Peters, Parasitology 1985, 90, 705.
- [2] A.R. Butler, Y. Wu, Chem. Soc. Rev. 1992, 85; D.L. Klayman, Science 1985, 228, 1049.
- [3] T.T. Hien, N.J. White, Lancet 1993, 341, 603; G.-S. Ding, Int. J. Exp. Clin. Chemother. 1988, 1, 9.
- [4] S.S. Zaman, R.P. Sharma, *Heterocycles* 1991, 32, 1593; China Cooperative Research Group on Qinghaosu and its Derivatives as Antimalarials, J. Tradit. Chim. Med. 1982, 2, 9 and 45.
- [5] Qinghaosu Antimalarial Coordinating Research Group, Chin. Med. J. 1979, 92, 811.
- [6] C. W. Jefford, E. C. McGoran, J. Boukouvalas, G. Richardson, B. L. Robinson, W. Peters, *Helv. Chim. Acta* 1988, 71, 1805.
- [7] C.W. Jefford, Y. Wang, G. Bernardinelli, Helv. Chim. Acta 1988, 71, 2042.
- [8] C. Earnshaw, C.J. Wallis, S. Warren, J. Chem. Soc., Perkin Trans. 1 1979, 3099.
- [9] F. Johnson, Chem. Rev. 1968, 68, 375.
- [10] W. Chatani, S. Fujii, Y. Yamasaki, S. Murai, N. Sonoda, J. Am. Chem. Soc. 1986, 108, 7361.
- [11] H.O. Kalinowski, S. Berger, S. Braun, 'Carbon-13 NMR Spectroscopy', J. Wiley & Sons, New York, 1988, Chap. 3.
- [12] Institute of Biophysics, Academia Sinica, Sci. Sin. 1979, 22, 1114.
- [13] J. B. Hendrickson, J. Am. Chem. Soc. 1967, 89, 7047.
- [14] V. Rautenstrauch, W. Thommen, K. H. Schulte-Elte, Helv. Chim. Acta 1986, 69, 1638; G. Rousseau, P. Le Perchec, J. M. Conia, Synthesis 1978, 67; D. Lerdal, C. S. Footc, Tetrahedron Lett. 1978, 3227.
- [15] R. E. Desjardins, C. J. Canfield, D. E. Haynes, J. D. Chulay, Antimicrob. Agents Chemother. 1979, 16, 710;
 W. K. Milhous, N. F. Weatherly, J. H. Bowdre, R. E. Desjardins, *ibid.* 1985, 27, 525.
- [16] M. J. O'Neill, D. H. Bray, P. Boardman, J. D. Phillipson, D. C. Warhurst, 'Antimalarial Drugs', Part 1, Planta Med. 1985, 5, 394.
- [17] S. Thaithong, G. H. Beale, Trans. R. Soc. Trop. Med. Hyg. 1981, 75, 271.
- [18] P. Nguyen-Dinh, D. Payne, Bull. WHO 1980, 58, 909.
- [19] C. W. Jefford, J. A. Velarde, G. Bernardinelli, *Tetrahedron Lett* 1989, 30, 4485; J. A. Velarde, Doctoral Thesis (No. 2358), University of Geneva, Switzerland, 1989.
- [20] G. H. Posner, C. H. Oh, L. Gerena, W. K. Milhous, J. Med. Chem. 1992, 35, 2459.
- [21] M. A. Avery, W. K. M. Chong, J. E. Bupp, J. Chem. Soc., Chem. Commun. 1990, 1487.
- [22] W. Peters, B. L. Robinson, J. C. Rossier, C. W. Jefford, Ann. Trop. Med. Parasitol. 1993, 87, 1; W. Peters, B. L. Robinson, J. C. Rossier, D. Misra, C. W. Jefford, *ibid*. 1993, 87, 9; W. Peters, B. L. Robinson, G. Tovey, J. C. Rossier, C. W. Jefford, *ibid*. 1993, 87, 111.
- [23] G. Stork, A. Brizzolara, H. Landesman, J. Szmuszkovicz, R. Terrell, J. Am. Chem. Soc. 1963, 85, 207.
- [24] W.C. Still, M. Kahn, A. Mitra, J. Org. Chem. 1978, 43, 2923.
- [25] M. Larchevêque, G. Valette, T. Cuvigny, Synthesis 1977, 424.
- [26] P. Main, S.J. Fiske, S.E. Hull, L. Lessinger, G. Germain, J.-P. Declercq, M.M. Woolfson, 'A System of Computer Programs for the Automatic Solution of Crystal Structures from X-Ray Diffraction Data', Universities of York, England, and Louvain-la-Neuve, Belgium, 1980.
- [27] J. M. Stewart, P. A. Machin, C. W. Dickinson, H. L. Ammon, H. Heck, H. Flack, 'The XRAY76 System, Tech. Rep. TR-446', Computer Science Center, University of Maryland, College Park, Maryland, 1976.
- [28] C. K. Johnson, 'ORTEP II; Report ORNL-5138', Oak Ridge National Laboratory, Oak Ridge, TN, 1976.

2788